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REVISED CLASSIFICATION AND PHYLOGENETIC HYPOTHESIS FOR THE ACANTHOSTOMINAE LOOSS, 1899 (DIGENEA: OPISTHORCHIFORMES: CRYPTOGONIMIDAE)

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Abstract. - Specimens of an acanthostome digenean originally identified as 5 Acanthostomum scyphocephalum and later transferred to Timoniella are described and named as a distinct species of Timoniella. Acanthostomum scyphocephalum sensu strictu is included in an updated phylogenetic analysis of the acanthostome digeneans. The new analysis differs from an earlier one by Brooks (1980) by allowing reversals (Wagner criterion vs. Camin-Sokal criterion), producing a more parsimonious representation of character data; no transformation series needed re-polarization. Acanthostomum scyphocephalum is a member of the clade containing all other species of Acanthostomum occurring in North, Central and South America. Acanthostomum is paraphyletic if Atrophecaecum is excluded from it; accordingly, the two genera are synonymized. No other changes from the hypothesis of Brooks (1980) resulted. The resulting annotated phylogenetic classification, with synapomorphic diagnoses, includes Acanthostomum as the sister-group of Caimanicola, Proctocaecum as their sister-group, the monotypic Gymnatrema as their sister-group, and Timoniella as the basal sister-group. Four new subgenera are proposed, one in Timoniella, one in Proctocaecum, and two in Acanthostomum.

The acanthostome digeneans (Opisthorchiformes: Cryptogonimidae: Acanthostominae) inhabit a variety of piscivorous poikilotherm amniote vertebrates throughout the tropical and subtropical regions of the world. Brooks (1980) provided the first phylogenetic systematic analysis of the acanthostomes. He recognized six genera, Timoniella Rebecq, 1960, Proctocaecum Baugh, 1957, Gymnatrema Morozov, 1955, Caimanicola Teixeira de Freitas & Lent, 1938, Acanthostomum Looss, 1899 and Atrophecaecum Bhalerao, 1940, although Acanthostomum had no synapomorphy to support its recognition as a monophyletic group.

Since that time, publications have described two additional species and discussed their phylogenetic relationships (Brooks & Caira 1982, Blair et al. 1988), and have added data about host and geographic distributions, as well as valuable taxonomic information (Ostrowski de Nuñez 1984a, 1984b, 1986, 1987). This study provides a description of a new species, and incorporates the new data provided by Ostrowski de Nuñez into the phylogenetic data base for the acanthostomes, resulting in an updated phylogenetic hypothesis and an annotated classification with cladistic diagnoses for all taxa.

Methods

In addition to the material specified in Brooks (1980), Brooks & Caira (1982) and Blair et al. (1988), we have examined the following material loaned from Dr. Margarita Ostrowski de Nuñez: *Acanthostomum gnerii* (19 specimens from *Rhamdia* sapo, Laguna Chis-Chil, Prov. Buenos Aires, Argentina); Acanthostomum megacetabulum (1 specimen from Drymarchon corais melanurus, Villahermosa, Mexico); Caimanicola marajoarum (14 specimens from Crocodvlus intermedius, Caracas, Venezuela: 3 specimens from Paleosuchus sp., Antioquia, Colombia); Caimanicola brauni (13 specimens from Phrynops hilarii, Buenos Aires, Argentina); Timoniella loossi (3 specimens from Crocodylus intermedius, Caracas, Venezuela); Acanthostomum sp. VI of Ostrowksi de Nuñez (1984b) (2 specimens from Caiman fuscus, Rio Chagras, Panama). All measurements are in µm unless otherwise noted. TBL = total bodylength. Figures were drawn with the aid of a drawing tube.

Results and Discussion

Timoniella ostrowskiae, new species Figs. 1-3

Synonyms. — Acanthostomum scyphocephalum of Mañe-Garzon & Gil, 1961; Timoniella scyphocephala of Brooks, 1980.

Description. - (based on holotype and 2 paratypes) Body 2.98-3.15 mm long by 420-540 wide at midbody. Tegument covered with spines of uniform size. Oral sucker terminal, bell-shaped, 410-517 long by 443-517 wide, armed with single row of 23 spines 68-78 long by 20-25 wide. Pharynx 197-205 long by 187-230 wide. Prepharynx 377-426 long. Ratio of oral sucker width to pharyngeal width 1:0.42-0.45. Ceca opening separately at posterior end of body. Acetabulum 1.22-1.47 mm from anterior end; forebody 41-47% TBL. Acetabulum 140-156 long by 131-147 wide; ratio of oral sucker width to acetabulum width 1:0.27-0.32. Testes tandem, intercaecal, near posterior end of body; posttesticular space 5.2-5.5% TBL. Anterior testis 187-205 long by 123-139 wide, posterior testis 180-221 long by 115-123 wide. Male genitalia consisting of coiled external seminal vesicle lying posterodorsal to acetabulum and musculoglandular pars prostatica and ejaculatory duct extending anteriorly dorsal to acetabulum, opening into genital atrium. Gonotyl lacking. Genital pore ventral, medial, immediately preacetabular. Ovary 90-139 anterior to anterior margin of anterior testis, 123-139 long by 115-123 wide. Seminal receptacle posterodorsal to ovary, between ovary and anterior testis. Mehlis gland present, Laurer's canal short. Uterine loops extending posteriorly to lateral margin of anterior testis, anteriorly to posterior margin of seminal vesicle, occupying 38-43% TBL; terminal portion of uterus opening into genital atrium. Vitellaria follicular, in two longitudinal extracecal fields extending from level of posterior margin of seminal vesicle to slightly posterior to anterior margin of anterior testis. Eggs 25-27 long by 10-12 wide.

Type host.—*Phrynops hilarii* (Dumeril and Bibron).

Type locality.—Rio Negro, Paso de los Toros, Departamento de Tacuarembo, Uruguay.

Holotype. – URFC Helm. Coll. No. 11038. Paratypes: URFC Helm. Coll. No. 11039–11040.

Etymology.—The species is named in honor of Dr. Margarita Ostrowski de Nuñez, who first recognized its distinct identity.

Brooks (1980) was unable to locate the holotype of A. scyphocephalum Braun, 1899 which had been collected in "Testudo matamata" from southern Brazil. Based on examination of specimens collected in Phrynops hilarii from Uruguay (reported by Mañe-Garzon & Gil 1961), Brooks (1980) transferred the species to Timoniella because it possessed preovarian rather than postovarian seminal receptacles, a unique and unreversed synapomorphy that diagnoses Timoniella among the acanthostomes. Ostrowski de Nuñez (1986) found the holotype of Acanthostomum scyphocephalum Braun, 1899, and showed that it was a member of Acanthostomum. The



Figs. 1-3. Timoniella (Maillardiella) ostrowskiae. 1. Ventral view of holotype. 2. Ootype region. 3. Terminal genitalia.

Table 1.—Homoplasious changes for 17 characters of acanthostomes. Character consistency index = number of apomorphic states for each character divided by total number of changes postulated on the phylogenetic trees. For character identities, see Appendix 1.

Char- acter state	Figure #-Character number on figure	Character consistency index
1-0	9-36	50% (1/2)
2-0	9-38	50% (1/2)
6-0	5-3	50% (1/2)
7-0	4-17, 5-12	50% (1/2)
9-0	8-10	
9-1	4-10, 5-1, 6-3, 7-5, 8-7, 9-32	33% (3/9)
10-0	6-1	
10-1	5-2, 7-4, 8-2, 8-17, 9-29, 9-35	17% (1/7)
11-1	5-4, 9-21	50% (1/2)
12-1	5-5, 6-11, 8-14	33% (1/3)
13-1	5-6, 6-6, 8-13, 9-22, 9-33	33% (2/6)
14-2	5-13, 6-7, 8-1	
14-3	5-14, 9-19	
14-5	4-12, 6-4, 8-6, 9-26	57% (8/14)
15-1	5-8, 8-18, 9-24	50% (2/4)
16-0	7-3	
16-1	4-20, 5-9, 6-8, 9-20, 9-28, 9-34	17% (1/7)
17-1	5-10, 8-3	67% (2/3)
18-0	8-30	
18-1	5-11, 7-1, 8-9	40% (2/5)
22-0	7-2, 8-11	50% (2/4)
23-1	6-5, 6-13, 9-25	33% (1/3)
29-1	5-15, 6-12, 8-15	50% (2/4)

specimens reported by Mañe-Garzon & Gil (1961) therefore appear to represent a previously unnamed species of Timoniella, which we formally described and named above. Timoniella ostrowskiae is most closely related to T. incognita Brooks, 1980, T. loossi (Perez Vigueras, 1957) Brooks, 1980, T. absita Blair et al., 1988, and T. unami (Pelaez & Cruz, 1957) Brooks, 1980, all of which lack gonotyls. Of those species, T. ostrowskiae is the only one exhibiting ceca opening separately at the posterior end of the body; T. incognita has blindly-ending ceca, and the remaining three species have ceca opening into the excretory vesicle near the posterior end of the body.

As indicated in the introduction, new information concerning acanthostomes has accumulated since Brooks (1980) produced

the first phylogenetic systematic hypothesis for the group. Brooks (1980) used the Camin-Sokal (also known as the Weighted Invariant Step Strategy [WISS]-Wiley et al. 1991) criterion, allowing no evolutionary reversals, in producing a phylogenetic tree for the acanthostomes. He found no synapomorphy to support the monophyly of Acanthostomum, nor any character that could link Acanthostomum with either Caimanicola or Atrophecaecum, two genera having synapomorphic support that were shown forming an unresolved trichotomy with Acanthostomum. In this study we reanalyzed the data using the less restrictive Wagner criterion (Wiley et al. 1991) after checking character polarizations using estimates of higher-level digenean phylogenetic relationships (Brooks et al. 1985, Brooks et al. 1989, Brooks & McLennan 1993) not available to Brooks (1980). We found support for the original character polarizations of Brooks (1980, see also Brooks & Caira 1982, Blair et al. 1988) but, as would be expected, we also found more parsimonious optimizations for some homoplasious characters when reversals were allowed (for a summary of transformation series, see Appendix 1; for a summary of homoplasious changes, including reversals, see Table 1). Based on the information presented by Ostrowski de Nuñez (1986), we included Acanthostomum scyphocephalum in the phylogenetic framework; it appears to be a member of the clade containing all the other species of Acanthostomum occurring in North, Central and South America (Fig. 8). Most importantly, the new analysis showed that Acanthostomum is paraphyletic (Fig. 8), and should be combined with Atrophecaecum (Figs. 8, 9).

Systematic theorists have begun investigating the problems of providing robust means for assessing the results of phylogenetic analyses (e.g., Archie 1989, Farris 1989, Sanderson & Donoghue 1989, Klassen et al. 1991, Meier et al. 1991). These studies have produced some interesting, and in some cases initially counter-intuitive, findings. For example, the minimum significant value for the most commonly used indicator, the consistency index (CI-Wiley et al. 1991), drops as one adds taxa and characters to a study; for example, a study using 50 characters for 20 taxa and reporting a CI of 65% may actually be more robust than using 10 characters for 7 taxa and reporting a CI of 80%. This happens because there are often apomorphic character changes occurring once within a given taxon that also occur once in another taxon. If the scope of a study were expanded to include both taxa, the estimate of homoplasy would increase (and the CI would drop) even if the hypothesized phylogenetic relationships of the (now) subgroups did not change. Or, to use current terminology, we would say that a global phylogenetic analysis had discovered homoplasy that the two local analyses failed to recognize. In some cases such global homoplasy could affect the hypotheses of relationships, so recognizing global homoplasy may play an important role in determining robust character polarities during outgroup comparisons at the inception of a phylogenetic analysis (e.g., Maddison et al. 1984, Wiley et al. 1991).

The acanthostomes provide an excellent illustration of the relationship between local and global parsimony considerations in phylogenetic analysis. If we treat all the acanthostomes as a single taxon, the consistency index for the characters reported herein is 50.5% (49 apomorphic character states and 97 character changes). This value is low for digeneans in general, the consistency index based on global parsimony considerations for all digeneans being approximately 72% (Brooks & McLennan 1993). By contrast, if we treat each of the clades denoted as a genus separately, the estimated homoplasy is generally much lower (100% for the characters used at the generic level by themselves, including the monotypic Gymnatrema, for the species of Timoniella, and for the species of Caimanicola, and 84.6% for the species of *Proctocaecum*), the exception being *Acanthostomum*, for which the consistency index based on local parsimony considerations is 51%. This means that although there is much homoplasy among the acanthostomes as a whole, most of it is dispersed among clades rather than concentrated within clades. Thus, determination of plesiomorphic states by outgroup comparisons is not problematical, and there is presently a single most parsimonious tree, for the group.

Mensural, or continuous variable, characters are problematical for phylogenetic reconstruction. The determination of discrete character states is often problematical for such traits; options seem to range from considering almost every species autapomorphic to recognizing very few states and there is no consensus approach among phylogeneticists. Brooks (1980) adopted a conservative approach to recognizing character states based on his examination of available specimens. Taking a conservative approach to such traits often results in considerable homoplasy. In the present study, the homoplasy is distributed among half (17 of 34)of the transformation series (1, 2, 6, 7, 9-18, 22-23, and 29 in Appendix 1), 9 of which (10, 12-13, 15-18, 22-23) are mensural in nature. Of the 48 homoplasious character transformations, 28 (58.3%) stem from the mensural characters and 20 (41.7%) from the qualitative traits; moreover, the mensural traits that show homoplasy have a combined character consistency index of 28.6% (14/42), while the qualitative traits that show homoplasy have a combined character consistency index of 48.7% (19/ 39) (Table 1). While suggestive, these data are actually moot with respect to the question of whether or not such characters provide adequate phylogenetic information, because they exhibit high levels of homoplasy but do not support relationships that are contradicted by non-mensural characters. A strong test of these characters requires a search for intrinsically qualitative

characters whose apomorphic states support phylogenetic relationships that conflict with the ones supported by the present data base.

Conclusions

The changes discussed above are reflected in the phylogenetic trees, cladistic diagnoses, and classification below (the monotypic Gymnatrema has no separate cladogram). In the following, italicized numbers refer to synapomorphies listed by number on Fig. 4; other numbers refer to synapomorphies listed by number on Figs. 5-8. Four new subgenera are proposed. Each is named for a digenean systematist who has advanced our understanding of acanthostome relationships: Dr. Claude Maillard, University of Montpellier, France; Dr. David Gibson, British Museum (Natural History), London, England; Dr. David Blair, James Cook University, Townsville, Australia; and Dr. Robin Overstreet, Gulf Coast Marine Research Laboratory, Ocean Springs, Mississippi, USA.

Subfamily Acanthostominae Poche, 1926 (Fig. 4)

Diagnosis.—Cryptogonimidae with terminal oral sucker (1); armed with single row of spines (2); preacetabular pit (3); genital pore not in preacetabular pit (4); seminal vesicle coiled posteriorly (5); suckerlike gonotyl present (6).

Genus Timoniella Rebecq, 1960 (Fig. 5)

Diagnosis.—Acanthostominae preovarian seminal receptacle (7).

Subgenus Timoniella Rebecq, 1960

Diagnosis. – Timoniella having vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (1); length of body occupied by uterine loops more than 50% TBL* (2); seminal vesicle not coiled posteriorly* (3); prepharynx shorter than pharynx* (4); ratio of oral sucker width to pharyngeal width 1:0.25-0.40* (5); ratio of body length to width averaging 7.5-15:1*(6).

T. praeterita (Looss, 1901) Maillard, 1974

Diagnosis.—Cyclocoel (7); forebody 10–20% TBL* (8); maximum body length 7–16 mm* (9).

T. imbutiformis (Molin, 1859) Brooks, 1980

Diagnosis.—Ratio of oral sucker : acetabular width 1:0.8–1.3* (10); oral spines averaging 25–30 in number* (11).

Subgenus Maillardiella, new subgenus

Diagnosis. – Timoniella lacking gonotyls* (12).

T. incognita Brooks, 1980

Diagnosis.—With characters of the subgenus.

Remarks: This species was originally reported by Nasir (1974) as *Acanthostomum scyphocephalum* inhabiting *Caiman crocodilus crocodilus*. According to Ostrowski de Nuñez (1984b), the acanthostomes reported as *Acanthostomum scyphocephalum* from *Drymarchon corais* by Nasir (1974) may be a still undescribed species.

T. ostrowskiae Brooks & Holcman, 1993

Diagnosis. - Ceca opening separately at posterior end of body* (13).

T. unami (Pelaez & Cruz, 1957) Brooks, 1980

Diagnosis.—Ceca opening into excretory vesicle* (14); vitelline follicles extending posteriorly to middle of posterior testis*(15).

T. loossi (Perez Vigueras, 1957) Brooks, 1980

Diagnosis.—Ceca opening into excretory vesicle* (14); vitelline follicles extending

posteriorly to middle of posterior testis* (15); vitelline follicles confluent dorsally (16).

T. absita Blair et al., 1988

Diagnosis.—Ceca opening into excretory vesicle* (14); vitelline follicles extending posteriorly to middle of posterior testis* (15); constriction in seminal vesicle (17).

Genus Gymnatrema Morozov, 1955

Diagnosis. — Acanthostominae having some uterine loops lateral to testes but none posttesticular (8); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (10); vitelline follicles confluent posttesticularly (11); one cecum atrophied* (12); one cecum opening laterally and one cecum ending blindly (13).

G. gymnarchi (Dollfus, 1950) Morozov, 1955

Diagnosis. – With characters of the genus.

Genus Proctocaecum Baugh, 1957 (Fig. 6)

Diagnosis. – Acanthostominae having some uterine loops lateral to testes but none posttesticular (8); ceca opening separately and laterally at even levels (9); excretory vesicle Y-shaped with short stem and constriction of arms in middle (14); eggs averaging more than 30 μ m long (15); gonotyl large, solid-muscular (16).

Subgenus Proctocaecum Baugh, 1957

Diagnosis.—*Proctocaecum* having relative length of uterine loops less than 45% TBL* (1).

P. gonotyl (Dollfus, 1950) Brooks, 1980

Diagnosis.—With characters of the subgenus.

P. vicinum (Odhner, 1902) Brooks, 1980

Diagnosis.—Ceca opening separately and laterally at uneven levels (2).

P. coronarium (Cobbold, 1861) Brooks, 1980

Diagnosis. —Ceca opening separately and laterally at uneven levels (2); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (3); one cecum atrophied* (4); maximum oral spine length more than 100 μ m* (5).

Subgenus Overstreetium, new subgenus

Diagnosis.—*Proctocaecum* having ratio of body length to width averaging 7.5-15: 1* (6).

P. productum (Odhner, 1902) Brooks, 1980

Diagnosis.—With characters of the subgenus.

 P. elongatum (Tubangui & Masiluñgen, 1936) Brooks, 1980

Diagnosis.—Ceca opening separately at posterior end of body* (7); maximum body length 7–16 mm* (8); ratio of body length to width averaging more than 20:1 (9).

P. crocodili (Yamaguti, 1954) Brooks, 1980

Diagnosis.—Ceca opening separately at posterior end of body* (7); maximum body length 7–16 mm* (8); forebody less than 10% TBL (10).

P. atae (Tubangui & Masiluñgen, 1936) Brooks, 1980

Diagnosis.—Ceca opening separately at posterior end of body* (7); maximum body length 7–16 mm* (8); ratio of oral sucker width to pharyngeal width 1:0.25-0.40*(11).

P. nicolli Brooks, 1980

Diagnosis.—Ceca opening separately at posterior end of body* (7); maximum body length 7–16 mm* (8); ratio of oral sucker width to pharyngeal width 1:0.25–0.40* (11); vitelline follicles extending anteriorly to posterior margin of acetabulum* (12); maximum oral spine length more than 100 μ m* (13).

Genus Caimanicola Teixeira de Freitas & Lent, 1938 (Fig. 7)

Diagnosis. — Acanthostominae having some uterine loops lateral to testes but none posttesticular (8); ceca opening separately and laterally at even levels (9); excretory vesicle Y-shaped with short stem and constriction of arms in middle (14); eggs averaging more than 30 μ m long (15); gonotyl lacking* (17); esophagus longer than pharynx (18); tegumental spines unusually robust in mid-forebody (19); maximum body length 7–16 mm* (20).

C. pavidus (Brooks & Overstreet, 1977) Brooks, 1980

Diagnosis.—Oral spines averaging 25–30 in number* (1).

C. caballeroi (Pelaez & Cruz, 1953) Brooks, 1980

Diagnosis. – Eggs averaging less than 30 μ m long (2); maximum body length 2–6 mm (3).

C. marajoarus Teixeira de Freitas & Lent, 1938

Diagnosis. – Eggs averaging less than 30 μ m long (2); length of body occupied by uterine loops more than 50% TBL* (4); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (5).

C. brauni (Mañe-Garzon & Gil, 1961) Brooks, 1980

Diagnosis. – Eggs averaging less than 30 μ m long (2); length of body occupied by uterine loops more than 50% TBL* (4); ratio of oral sucker width to acetabular width 1:0.3–0.7 (6).

Remarks: According to Ostrowksi de Nuñez (1984b), *Acanthostomum brauni* of Caballero (1955) is an undetermined species of acanthostome, possibly undescribed. Genus Acanthostomum Looss, 1899 (Figs. 8, 9)

Diagnosis. — Acanthostominae having some uterine loops lateral to testes but none posttesticular (8); ceca opening separately and laterally at even levels (9); eggs averaging more than 30 μ m long (15); gonotyl lacking* (17); excretory vesicle with long stem and short arms (21).

Subgenus Blairium, new subgenus

Diagnosis. — Acanthostomum having ceca opening separately at posterior end of body* (1).

A. scyphocephalum (Braun, 1899) Hughes et al., 1941

Diagnosis.—With characters of the subgenus.

Remarks: Ostrowski de Nuñez (1986) redescribed this species from the type material, which had been missing and presumed lost (see Brooks 1980).

A. americanum (Perez Vigueras, 1957) Herber, 1961

Diagnosis.—Length of body occupied by uterine loops more than 50% TBL* (2).

A. megacetabulum Thatcher, 1963

Diagnosis.—Length of body occupied by uterine loops more than 50% TBL* (2); ratio of oral sucker : acetabular width 1:0.8–1.3* (3).

A. gnerii Szidat, 1954

Diagnosis.—Testes oblique (4); vitelline follicles sparse (5).

A. minimum Stunkard, 1938

Diagnosis.—Testes oblique (4); vitelline follicles sparse (5); one cecum atrophied* (6).

A. astorquii Watson, 1976

Diagnosis. - Testes oblique (4); vitelline



Fig. 4. Phylogenetic relationships among the genera of the subfamily Acanthostominae. Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.

follicles sparse (5); one cecum atrophied* (6).

Subgenus Gibsonium, new subgenus

Diagnosis.—Acanthostomum having vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (7); oral spines averaging less than 20 in number* (8).

A. absconditum (Looss, 1901) Poche, 1926

Diagnosis. — With characters of the subgenus.



Fig. 5. Phylogenetic relationships among species of *Timoniella*. Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.



Fig. 6. Phylogenetic relationships among species of *Proctocaecum*. Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.

Subgenus Acanthostomum Looss, 1899

Diagnosis. — Acanthostomum having oral spines averaging 25–30 in number* (9); vitelline follicles extending anteriorly to posterior margin of seminal vesicle* (10); eggs averaging less than 30 μ m long (11); cecal bifurcation approximately 10% TBL preacetabular (12).

A. knobus Issa, 1962

Diagnosis.—Ratio of body length to width averaging 7.5-15:1*(13).

A. spiniceps (Looss, 1896) Looss, 1899

Diagnosis.—Ratio of oral sucker width to pharyngeal width 1:0.25–0.40* (14).



Fig. 7. Phylogenetic relationships among species of *Caimanicola*. Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.



Fig. 8. Phylogenetic relationships among species of *Acanthostomum* excluding *Acanthostomum* (*Atrophecaecum*). Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.

A. niloticum Issa, 1962

Diagnosis.—Ratio of oral sucker width to pharyngeal width 1:0.25–0.40* (14); vitelline follicles extending anteriorly to posterior margin of acetabulum* (15).

Subgenus Atrophecaecum Bhalerao, 1940

Diagnosis.—*Acanthostomum* having oral spines averaging 25–30 in number* (9); vitelline follicles extending anteriorly to pos-

terior margin of seminal vesicle* (10); vitelline follicles terminating preovarially (16); length of body occupied by uterine loops more than 50% TBL* (17); forebody 10– 20% TBL* (18).

A. indicum Sinha, 1942

Diagnosis. — With characters of the subgenus.

A. slusarskii Kalyankar, 1977



Fig. 9. Phylogenetic relationships among species of *Acanthostomum (Atrophecaecum)*. Numbers refer to apomorphic traits listed in diagnoses in text. Each asterisk (*) indicates the presence of a homoplasious character; the particular homoplasious characters are denoted by an asterisk in the diagnoses in the text.

Diagnosis.—Ceca opening into excretory vesicle* (19); maximum body length 7–16 mm* (20).

A. pakistanense Coil & Kuntz, 1960

Diagnosis. – Prepharynx shorter than pharynx* (21); forebody 10–20% TBL* (22); ratio of body length to width averaging 7.5– 15:1* (23); oral spines averaging 20–24 in number* (24); maximum oral spine length more than 100 μ m* (25).

A. asymmetricum (Simha, 1958) Khalil, 1963

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum lost (27); maximum body length 7–16 mm* (28).

A. proctophorum (Dwivedi, 1966) Yamaguti, 1971

Diagnosis. — Prepharynx shorter than pharynx* (21); one cecum lost (27); length of body occupied by uterine loops more than 50% TBL* (29); oral spines averaging 20– 24 in number* (30); vitelline follicles confluent preovarially (31).

A. simhai Khalil, 1963

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum atrophied* (26); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (32); ratio of body length to width averaging 7.5– 15:1* (33); maximum body length 7–16 mm* (34).

A. burminis (Bhalerao, 1926) Bhalerao, 1936

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum atrophied* (26); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (32); length of body occupied by uterine loops more than 50% TBL* (35).

A. lobacetabulare Brooks & Caira, 1982

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum atrophied* (26); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (32); subterminal mouth* (36); lobate acetabulum (37).

A. cerberi (Fischthal & Kuntz, 1965) Brooks, & Caira 1982

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum atrophied* (26); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (32); subterminal mouth* (36); no oral spines* (38); no esophagus (39).

A. marinum (Coil & Kuntz, 1960) Brooks & Caira, 1982

Diagnosis. – Prepharynx shorter than pharynx* (21); one cecum atrophied* (26); vitelline follicles not extending anteriorly to posterior margin of seminal vesicle* (32); subterminal mouth* (36); no oral spines* (38); no prepharynx (40); secondary group of vitelline follicles surrounding testes (41).

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Literature Cited

- Archie, J. W. 1989. Homoplasy excess ratios: new indices for measuring levels of homoplasy in phylogenetic systematics and a critique of the consistency index.—Systematic Zoology 38:253– 269.
- Baugh, S. C. 1957. Contributions to our knowledge of digenetic trematodes. II. – Proceedings of the National Academy of Science of India 26:295– 313.
- Bhalerao, G. 1926. On the trematode parasites of a water-snake, *Tropidonotus piscator*. – Parasitology 18:4–13.
 - ——. 1936. Studies on the helminths of India. Trematoda II.—Journal of Helminthology 14: 181–206.

 1940. Observations on the anatomy of Acanthostomum burminis (Bhalerao, 1926).—Indian Journal of Veterinary Science and Animal Husbandry 10:94–97.

- Blair, D., D. R. Brooks, J. Purdie, & L. Melville. 1988. *Timoniella absita* n. sp. (Digenea: Cryptogonimidae) from the saltwater crocodile (*Crocodylus porosus* Schneider) from Australia.—Canadian Journal of Zoology 66:1763–1766.
- Braun, M. 1899. Weitere Mitteilungen uber endoparasitische Trematoden der Chelonien. – Centralblatt fur Bakteriologie und Parasitenkunde 25:627–632.
- Brooks, D. R. 1980. Revision of the Acanthostominae (Digenea: Cryptogonimidae).—Zoological Journal of the Linnean Society 70:313-382.
 - —, & J. N. Caira. 1982. Atrophecaecum lobacetabulare sp. n. (Digenea: Cryptogonimidae: Acanthostominae) with discussion of the generic status of Paracanthostomum Fischthal and Kuntz, 1965 and Ateuchocephala Coil and Kuntz, 1960. – Proceedings of the Biological Society of Washington 95:223–231.
 - ——, & D. A. McLennan. 1993. Parascript: parasites and the language of evolution. Smithsonian Institution University Press, Washington, 429 pp.
 - —, & R. M. Overstreet. 1977. Acanthostome digeneans from the American Alligator in the southeastern United States.—International Journal of Parasitology 8:267–273.
 - , R. T. O'Grady, & D. R. Glen. 1985. Phylogenetic analysis of the Digenea (Platyhelminthes: (Cercomeria) with comments on their adaptive radiation.—Canadian Journal of Zoology 63:411–443.
 - —, S. M. Bandoni, C. A. Macdonald, & R. T. O'Grady. 1989. Aspects of the phylogeny of the Trematoda Rudolphi, 1808 (Platyhelminthes: Cercomeria).—Canadian Journal of Zoology 67:2609–2624.
- Caballero, E. 1955. Helmintos de la Republica de Panama. XVIII. Algunos trematodes de crocodilianos. 1a parte.—Anales de Instituto de Biologia de Mexico 26:433–446.
- Cobbold, T. S. 1861. List of Entozoa, including pentastomes, from animals dying at the Society's menagerie, between the years 1857–1860 inclusive with descriptions of several new species.— Proceedings of the Zoological Society of London 8:117–128.
- Coil, W. H., & R. E. Kuntz. 1960. Three new genera of trematodes from Pacific sea serpents. – Proceedings of the Helminthological Society of Washington 27:145–150.
- Dollfus, R. P. 1950. Trematodes recoltes au Congo belge par le Professeur Paul Brien (mai-aout 1937).—Annales de la Musee du Congo Belge C-Dierking 1:1–136.

- Dwivedi, M. P. 1966. On a new species of *Haplo-caecum* Simha, 1958 (Trematoda: Digenea) from a water snake *Tropidonotus piscator*. Revista Biologia Tropical 14:87–91.
- Farris, J. S. 1989. The retention index and the rescaled consistency index. - Cladistics 5:417-419.
- Fischthal, J. H., & R. E. Kuntz. 1965. Digenetic trematodes of amphibians and reptiles from North Borneo (Malaysia).—Proceedings of the Helminthological Society of Washington 32:124– 136.
- Herber, E. C. 1961. Some parasites from El Salvador.—Proceedings of the Pennsylvania Academy of Sciences 35:32–44.
- Hughes, R. C., J. W. Higginbotham, & J. W. Clary. 1941. The trematodes of reptiles. Part II, host catalogue. – Proceedings of the Oklahoma Academy of Sciences 21:35–43.
- Issa, G. I. 1962. Description of Acanthostomum niloticum n. sp. and Acanthostomum spiniceps knobus n. sub sp. (Trematoda, Acanthostomidae) from the river Nile, Egypt. – Wildlife Disease Association Micro-card No. 31.
- Kalyankar, S. D. 1977. Acanthostomum slusarskii sp. n. (Trematoda: Acanthostomidae) from Crocodilus palustris Less. in India.—Acta Parasitologica Polonica 24:227-230.
- Khalil, L. F. 1963. On Acanthostomum gymnarchi (Dollfus, 1950) with notes on the genera Acanthostomum Looss, 1899, Atrophecaecum Bhalerao, 1940, Gymnatotrema Morosov, 1955, and Haplocaecum Simha, 1958.—Journal of Helminthology 37:207–214.
- Klassen, G. J., R. D. Mooi, & A. Locke. 1991. Consistency indices and random data.—Systematic Zoology 40:446–457.
- Looss, A. 1896. Recherches sur la faune parasitaire de l'Egypte.-Memoires sur l'Egypte, Institut d'Egypte 3:1-252.
 - ——. 1899. Weitere Beitrage zur Kenntnis der Trematoden-Fauna Aegyptens, zugleich Versuch einer naturliches Gliederung des Genus Distomum Retzius.—Zoologische Jahrbuch, Abteilung fur Systematik 12:578–582.
 - —. 1901. Ueber die Fascioliden genera Stephanochasmus, Acanthochasmus, und einiger andere.—Zentralblatt fur Bakteriologie und Parasitenkunde 29:595–661.
- Maddison, W. P., M. J. Donoghue, & D. R. Maddison. 1984. Outgroup analysis and parsimony.—Systematic Zoology 33:83–103.
- Maillard, C. 1974. Cycle evolutif de *Timoniella praeteritum* (Looss, 1901) (Trematoda: Acanthostomidae) parasite de *Morone labrax* (Teleostei, Serranidae).—Bulletin de la Societe Zoologique de France 99:245–257.
- Mañe-Garzon, F., & O. Gil. 1961. Trematodos de las tortugas del Uruguay, II.-Communica-

ciones del Museo de Historia Natural de Montevideo 87:1-6.

- Meier, R., P. Kores, & S. Darwin. 1991. Homoplasy slope ratio: a better measurement of observed homoplasy in cladistic analyses.—Systematic Zoology 40:74–88.
- Molin, R. 1859. Nuovi myzelmintha raccolti ed esaminati. – Sitzungsberichte der Kaiserliche Academie der Wissenschaften, Wien, Mathematik-Naturwissenschaften Klasse 33:287–303.
- Morozov, F. N. 1955. Heterophyata. Vol. 10. Pp. 241–335 in K. I. Skrjabin, ed., Trematodes of animals and man. Academy of Sciences of the USSR, Moscow.
- Nasir, P. 1974. Revision of genera Acanthostomum Looss 1899 and Telorchis Luehe 1899 (Trematoda, Digenea) with redescription of Acanthostomum (Acanthostomum) scyphocephalum (Braun, 1901) and Telorchis aculeatus (von Linstow 1879) Braun 1901.—Rivista di Parassitologia 35:1-22.
- Odhner, T. 1902. Trematoden und reptilien nebst allgemeinen systematischen Bemerkungen.-Ofversigt af Konigliche Vetenskaps Akademiens Forhandlingar 59:19-45.
- Ostrowski de Nuñez, M. 1984a. Redescripcion de Acanthostomum marajoarum (Freitas y Lent, 1938) Hughes, Higginbotham and Clary, 1941 (Trematoda) sobre los ejemplares originales.— Physis 42:25-27.
- . 1984b. Beitrage zur Gattung Acanthostomum (Trematoda, Acanthostomidae) und zu den Entwicklungszyklen von A. marajoarum (Freitas & Lent, 1938) und A. loossi (Perez Vigueras, 1957) in Venezuela. – Mitteilungen der Zoologische Museum Berlin 60:179–201.
 - —. 1986. Acanthostomum scyphocephalum (Braun, 1899) Hughes, Higginbotham and Clary, 1941: Neubeschreibung des typischen materials aus dem Naturhistorischen Museum in Wien. – Annales der Naturhistorischen Museum Wien 87:331-337.
- . 1987. Der Entwicklungszyklus von Acanthostomum brauni Mane Garzon und Gil, 1961 (Trematoda, Acanthostomatidae). – Zoologisches Anzeiger 218:273–286.
- Pelaez, I., & F. Cruz. 1957. Consideraciones sobre el genero Acanthostomum Looss, 1899 (Trematoda: Acanthostomidae) con descripcion de dos especies de Mexico. — Memorias de la Congresa de las Ciencias Mexicana 7:269–284.
- Perez Vigueras, J. 1957. Contribucion al conocimiento de la fauna helmintologica cubana. – Memorias sobre la Historia Natural de la Isla de Cuba 23:1–36.
- Poche, F. 1926. Das System der Platoden. Archiven fur Naturgewissenschaften, Berlin, 458 pp.
- Rebecq, J. M. 1960. *Timoniella atherinae* nov. gen. nov. sp. (Trematoda: Acanthostomatidae) par-

asite d'*Atherina mochon* C.V.-Libro Homenaje Caballero y Caballero, Mexico, pp. 257-262.

- Sanderson, M. J., & M. J. Donoghue. 1989. Patterns of variation in levels of homoplasy. – Evolution 43:1781–1795.
- Simha, S.S. 1958. Studies on the trematode parasites of reptiles found in the Hyderabad state.—Zeitschrift fur Parasitenkunde 18:161–218.
- Sinha, B. B. 1942. Studies on the trematode parasites of reptiles. Part I. A new trematode, *Acanthostomum indicum*, belonging to the family Acanthostomidae, from the intestine of a crocodile.— Proceedings of the Indian Academy of Science 16:86–90.
- Stunkard, H. W. 1938. Further observations on the occurrence of anal openings in digenetic trematodes. – Zeitschrift fur Parasitenkunde 3:713– 725.
- Szidat, L. 1954. Trematodes nuevos de peces da agua dulce de la Republica Argentina.—Revista del Instituto de Investigacion de las Ciencias Naturales 3:1-85.
- Teixeira de Freitas, J. F., & H. Lent. 1938. Pequisas helminthologicas realisadas no Estada de Para.
 II. Dois novos trematodeos de *Caiman sclerops.* – Memorias do Instituto Oswaldo Cruz 33: 53–56.
- Thatcher, V. E. 1963. Studies on the trematodes of the Mexican Indigo Snake (Drymarchon corais melanurus) with descriptions of two new species.—Transactions of the American Microscopical Society 82:371–380.
- Tubangui, M. A., & V. Masiluñgen. 1936. Trematode parasites of Philippine vertebrates. VIII. Flukes from a cobra and a crocodile – Philippines Journal of Science 60:255–266.
- Watson, D. E. 1976. Digenea of fishes from Lake Nicaragua. Pp. 251–260 in T. B. Thorson, ed., Investigations of the ichthyofauna of Nicaraguan lakes. School of Life Sciences, University of Nebraska, Lincoln, 663 pp.
- Wiley, E. O., D. Siegel-Causey, D. R. Brooks, & V. A. Funk. 1991. The compleat cladist. Special Publication Number 19, University of Kansas Museum of Natural History Press, Lawrence, 158 pp.
- Yamaguti, S. 1954. Parasitic worms mainly from Celebes. Part 4. Trematodes of reptiles and birds.— Acta Medica Okayama 8:329–340.
- ——. 1971. Synopsis of digenetic trematodes of vertebrates. Keigaku Publishing Company, Tokyo, 1063 pp.

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Appendix 1

Transformation series for characters used to formulate phylogenetic hypotheses for acanthostome digeneans (for outgroup argumentation, see Brooks 1980, Brooks & Caira 1982, Brooks et al. 1985, Brooks et al. 1989). 0 = plesiomorphic; 1 or higher indicates apomorphic states. U = nonlinear transformation series, run unordered in computer-assisted analyses (numbers assigned to each apomorphic state are arbitrary).

- 1. Oral sucker subterminal (0); terminal (1).
- 2. Oral sucker lacking spines (0); armed with single row of spines (1).
- 3. Preacetabular pit lacking (0); present (1).
- 4. Ventrogenital pit present (0); lacking (1).
- 5. Genital pore in preacetabular pit (0); not in preacetabular pit (1).
- Seminal vesicle not coiled posteriorly (0); coiled posteriorly (1).
- Gonotyl lacking (0); suckerlike (1); large, solidmuscular (2). U
- 8. Seminal receptacle postovarian (0); preovarian (1).
- Vitelline follicles extending anteriorly to posterior margin of seminal vesicle (0); not extending anteriorly to posterior margin of seminal vesicle (1); confluent dorsally (2); confluent posttesticularly (3). U
- Space occupied by uterine loops less than 45% TBL (0); more than 50% TBL (1).
- Prepharynx longer than pharynx (0); shorter than pharynx (1).
- 12. Ratio of oral sucker width to pharyngeal width averaging 1:0.5 (0); 1:0.25–0.40 (1).
- 13. Ratio of body length to width averaging less than 7.5:1 (0); 7.5–15:1 (1); more than 20:1 (2). U
- Ceca ending blindly near posterior end of body (0); cyclocoel (1); opening separately at posterior end of body (2); opening into excretory vesicle (3); opening separately and laterally at even levels (4); one cecum atrophied (5); one cecum opening lat-

erally and one cecum ending blindly (6); opening separately and laterally at uneven levels (7); one cecum lost (8). U

- 15. Forebody more than 20% TBL (0); 10-20% TBL (1); less than 10% TBL (2).
- 16. Maximum body length less than 7 mm (0); 7–16 mm (1).
- 17. Ratio of oral sucker : acetabular width 1:0.6–0.9 (0); 1:0.8–1.3 (1); 1:0.3–0.7 (2).
- Oral spines averaging 20-24 in number (0); 25-30 (1); less than 20 (2). U
- 19. No constriction in seminal vesicle (0); constriction present (1).
- 20. Posttesticular loops present (0); some uterine loops lateral to testes but none posttesticular (1).
- Excretory vesicle Y-shaped with long stem (0); with short stem and constriction of arms in middle (1); long stem and short arms (2).
- 22. Eggs averaging less than 30 μ m long (0); more than 30 μ m long (1).
- 23. Maximum oral spine length less than $100 \,\mu\text{m}$ long (0); more than $100 \,\mu\text{m}$ (1).
- 24. Esophagus shorter than pharynx (0); longer than pharynx (1).
- 25. Tegumental spines not unusually robust in midforebody (0); unusually robust in mid-forebody (1).
- 26. Testes tandem (0); oblique (1).
- 27. Vitelline follicles numerous (0); sparse (1).
- 28. Cecal bifurcation averages 20% TBL preacetabular (0); 10% TBL preacetabular (1).
- 29. Vitelline follicles terminating at least at ovarian level and no further posterior than posterior margin of the ovary (0); extending posteriorly to middle of posterior testis (1); terminating preovarially (2). U
- 30. Vitelline follicles not confluent preovarially (0); confluent preovarially (1).
- 31. Acetabulum not lobate (0); lobate (1).
- 32. Esophagus present (0); lacking (1).
- 33. Prepharynx present (0); lacking (1).
- 34. Secondary group of vitelline follicles surrounding testes lacking (0); present (1).